

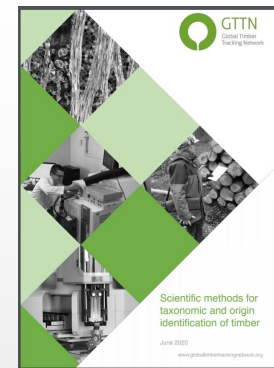
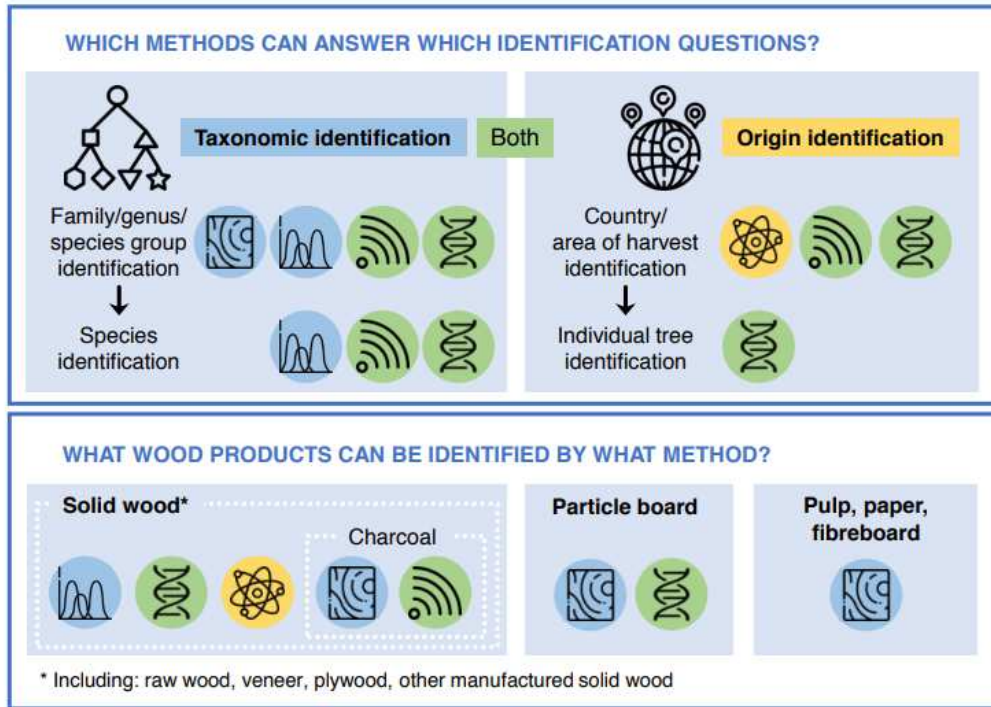
Tracing the geographical origin of timber with genetic analysis

Céline Blanc-Jolivet, Malte Mader, Niels Müller, Bernd Degen
Thünen Institute of Forest Genetics, Grosshansdorf, Germany



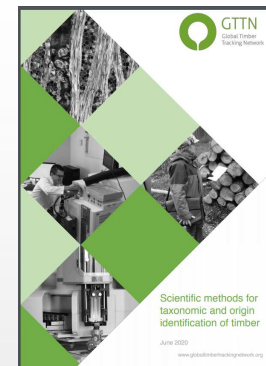
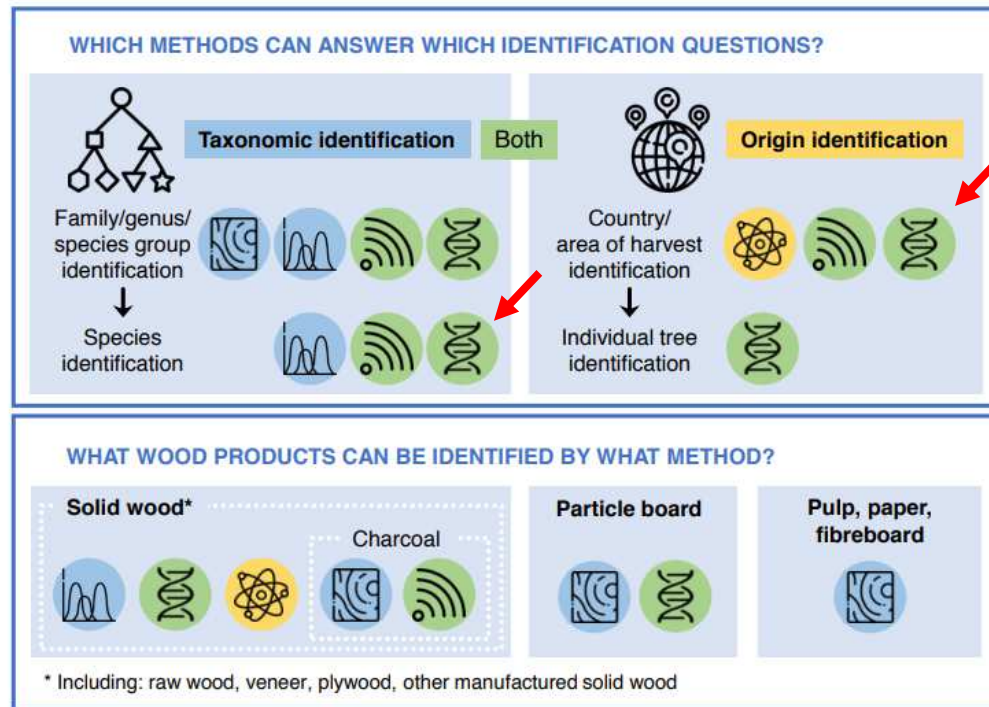
Which methods are available?

The question is... the question to answer !



Which methods are available?

The question is... the question to answer !



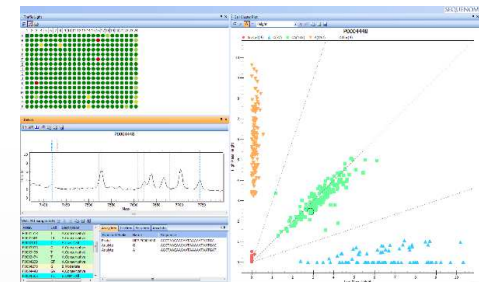
Which steps are necessary to apply the methods?

Development of reference data



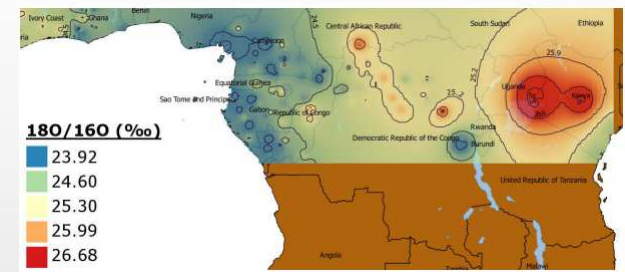
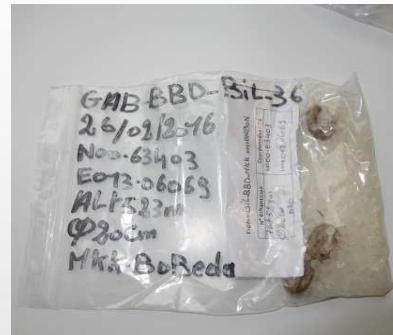
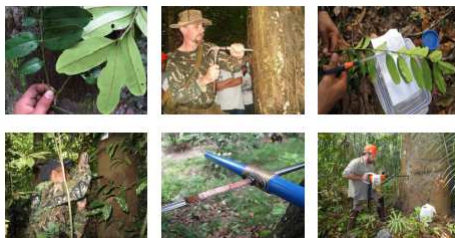
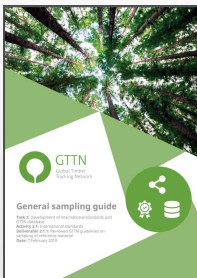
Dried Leaf/
cambium: **DNA**

Wood: **other
methods**



Georeferenced reference trees
with known species

Reference data

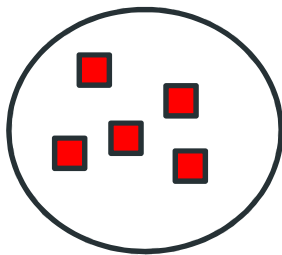


Genetic reference data ?

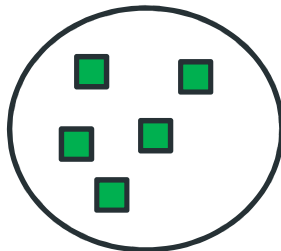
- **Important: the question is... the question to answer**
- **Geographical origin: which scale?**
 - Country?
 - Region?
 - Forest concession/stand?
- **The scale chosen defines the spatial resolution of the sampling and the analysis**

How do I get the appropriate genetic information?

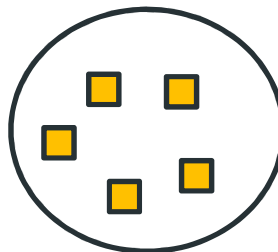
- The ideal case: the genotypes are “region specific”



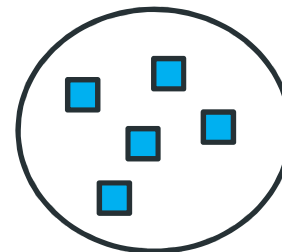
Country/Region 1



Country/Region 2

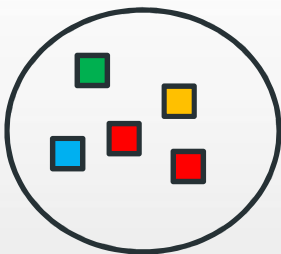


Country/Region 3

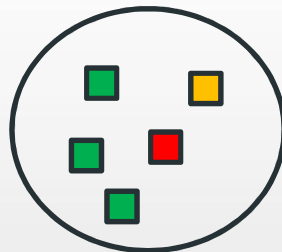


Country/Region 4

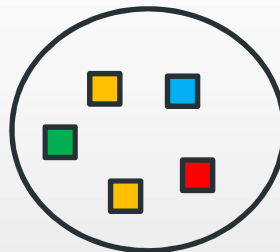
- This would likely happen with random genetic information:



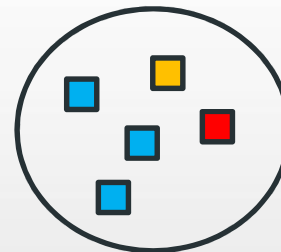
Country/Region 1



Country/Region 2



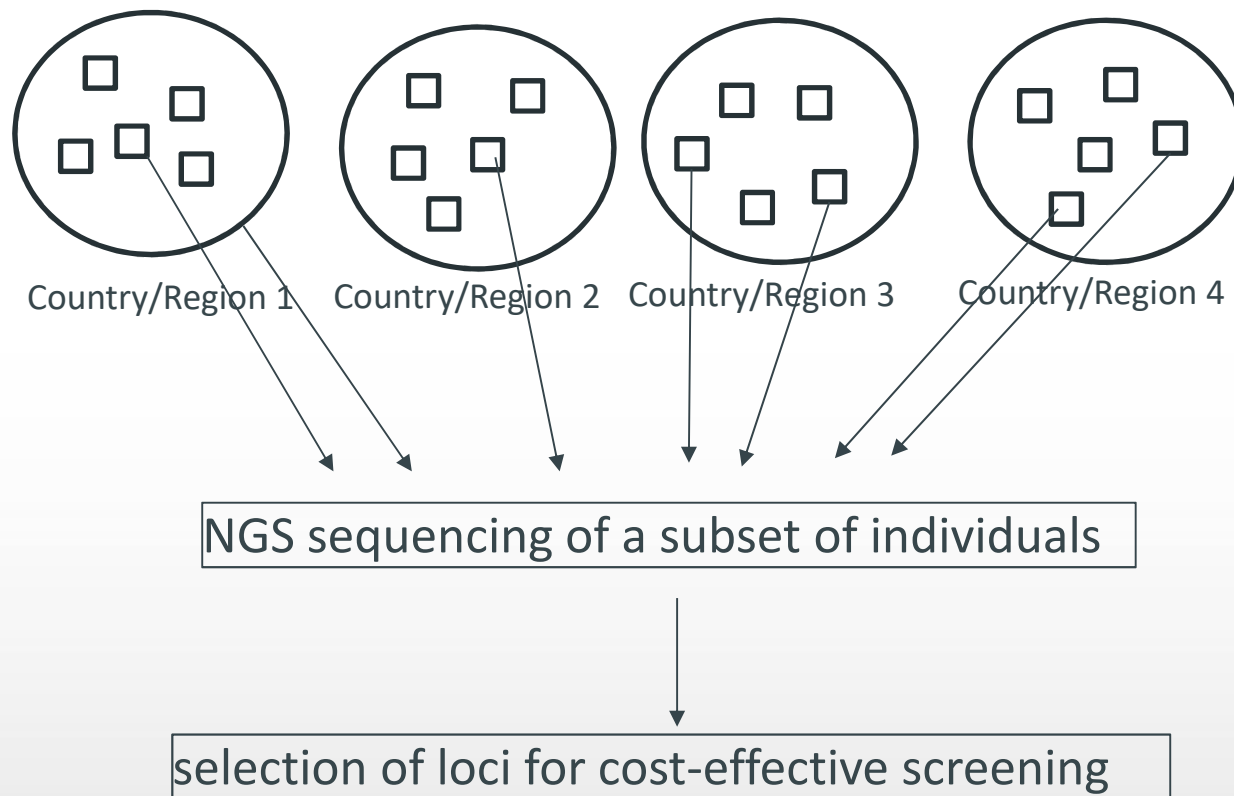
Country/Region 3



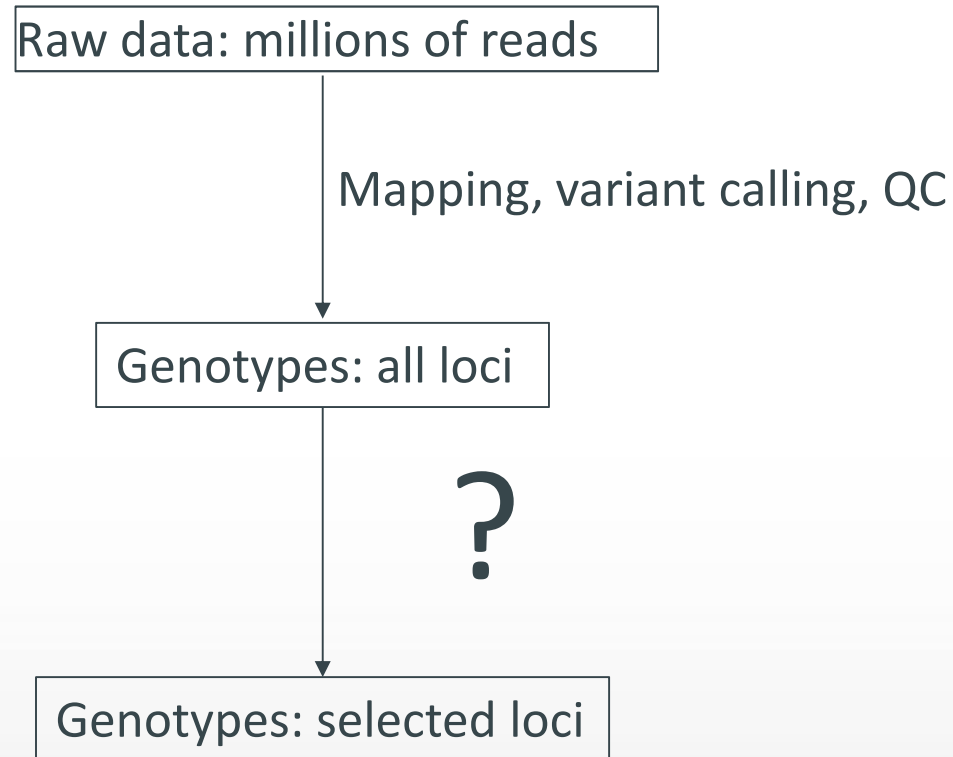
Country/Region 4

How do I get the appropriate genetic information?

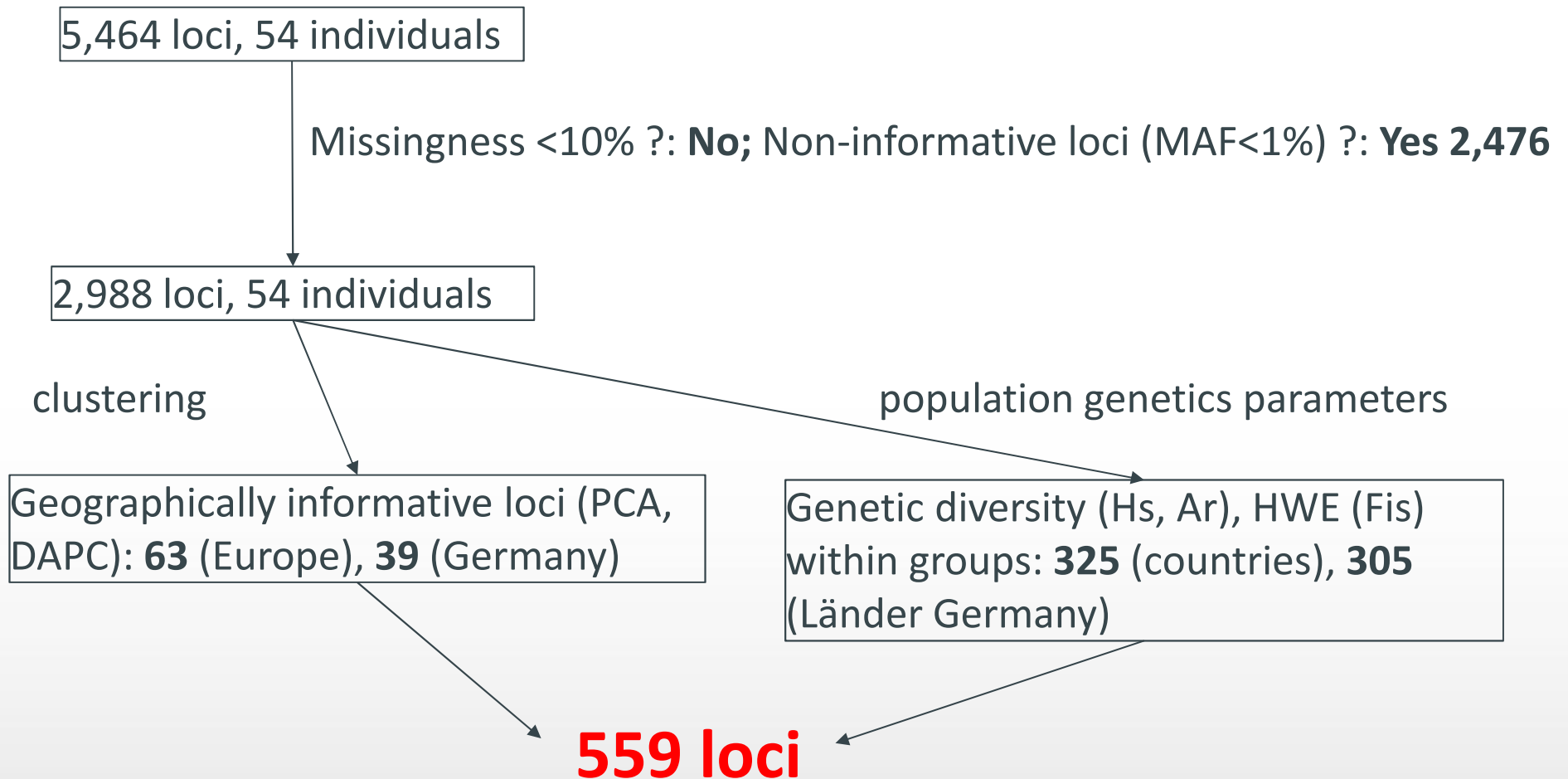
- How can I find the interesting loci among several millions?



Analysis of NGS data

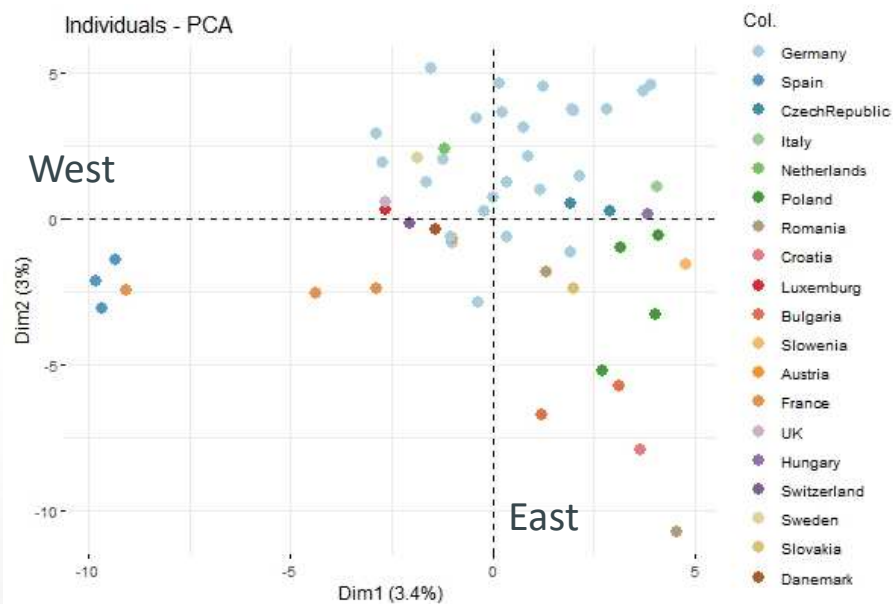


Locus selection: example ddRADseq data Beech

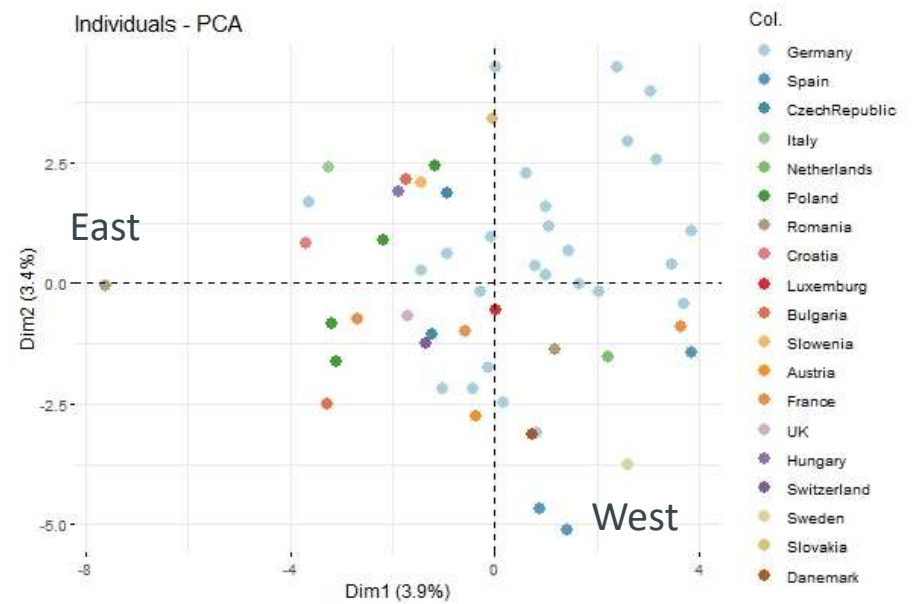


Locus selection: example ddRADseq data Beech

Is loss of information after marker selection acceptable?



5,464 loci, 54 individuals

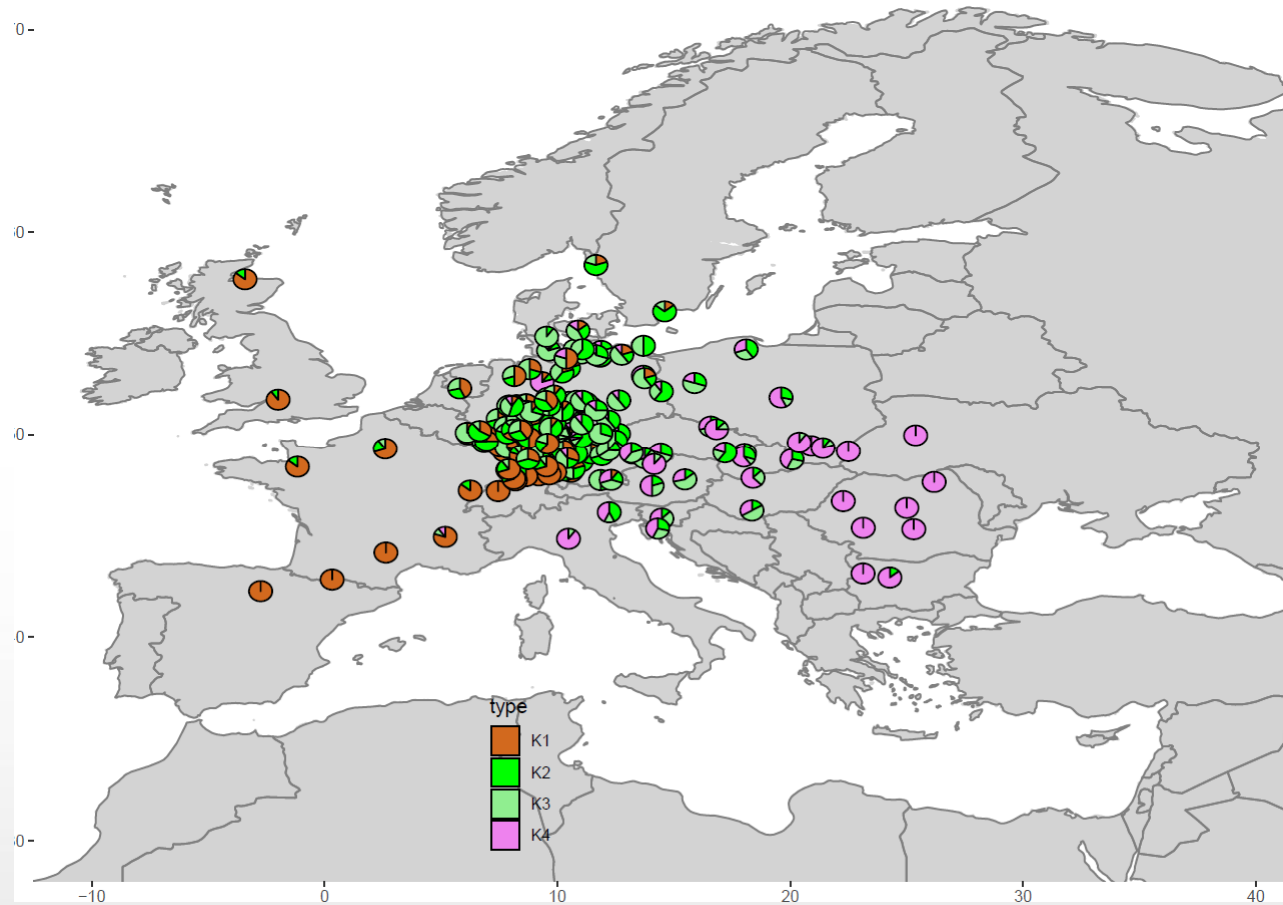


559 loci, 54 individuals

Reference data beech

- **2,000 individuals**
- **543 loci (SeqSNP method)**
- **Cleaning of data: 1,969 individuals @ 507 loci**
 - 31 individuals with >10 % missingness or duplicates
 - 10 loci with > 20 % missingness
 - 7 non-informative loci with rare alleles (MAF < 1%)
 - 19 loci with heterozygote excess or deficit

Reference data beech

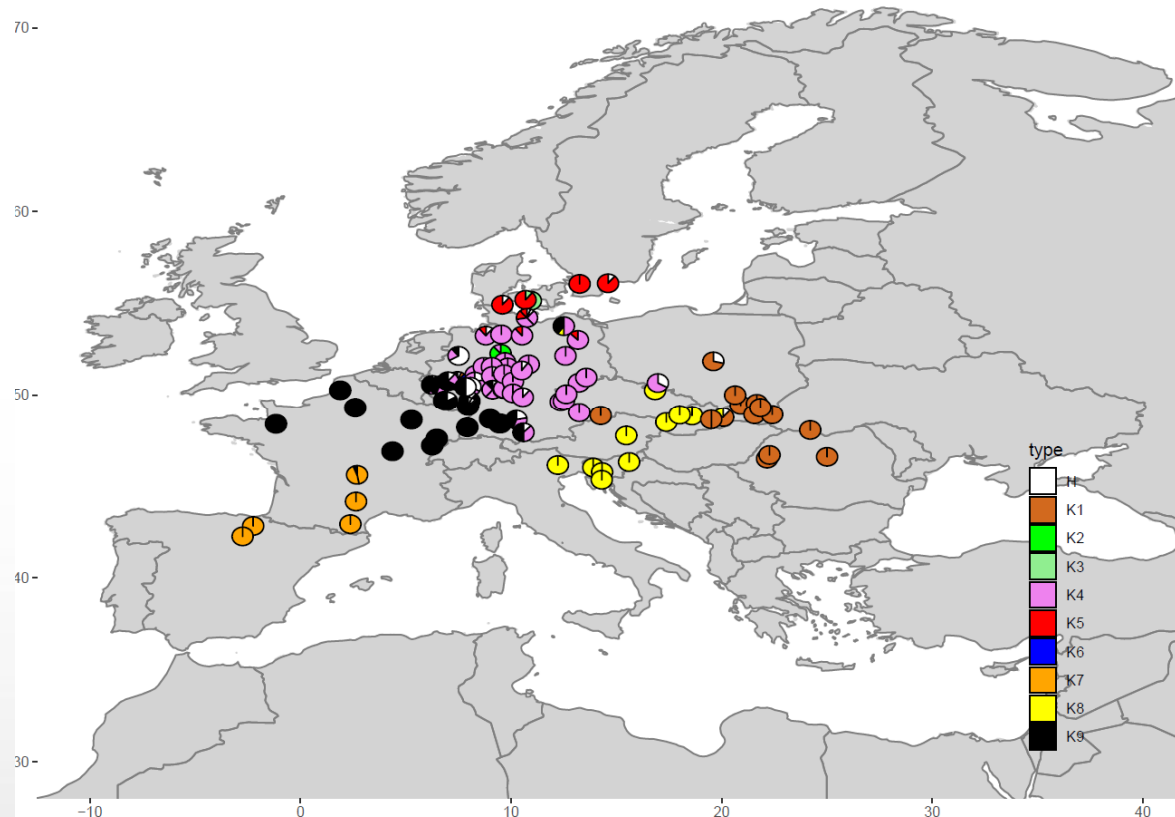


3 genetic groups

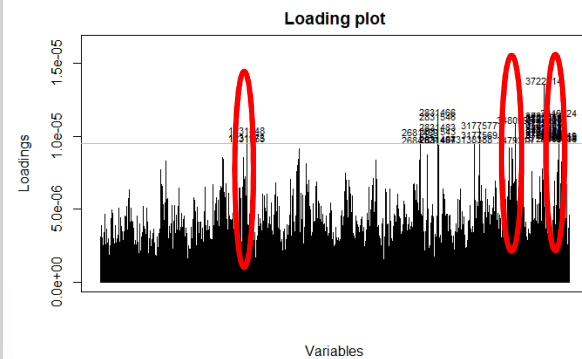
DAPC analysis

Whole-genome sequencing: higher resolution

860 individuals, 3.6 millions variants



DAPC analysis, 6 main groups

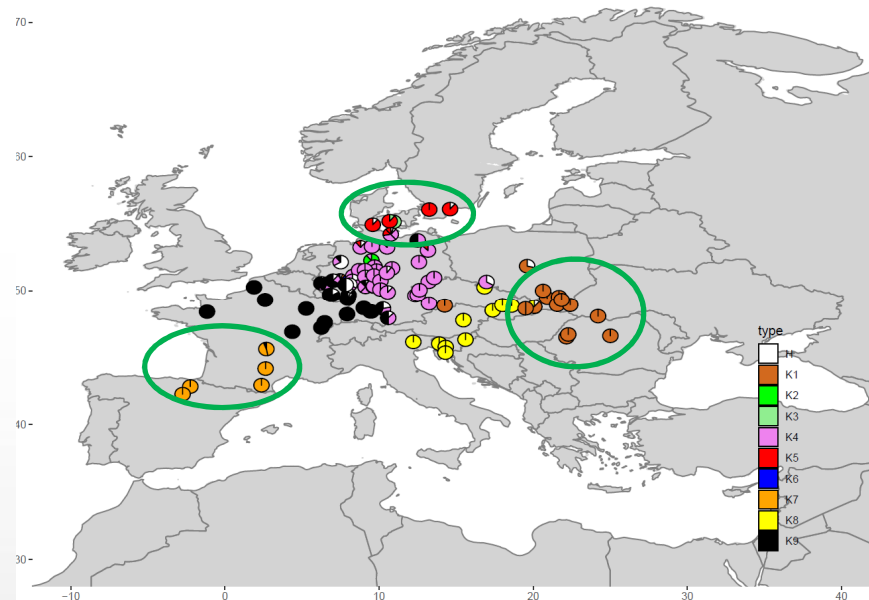


Top variants on first 3 axis: 92

Whole-genome sequencing: higher resolution

- From 92 variants 28 “non redundant” variants selected
- Performance of the marker set tested with self-assignment

Group	Correct	Total	% correct
K1	107	127	84.3
K4	116	307	37.8
K5	39	45	86.7
K7	42	51	82.4
K8	46	86	53.5
K9	108	171	63.2
Total	458	787	58.2

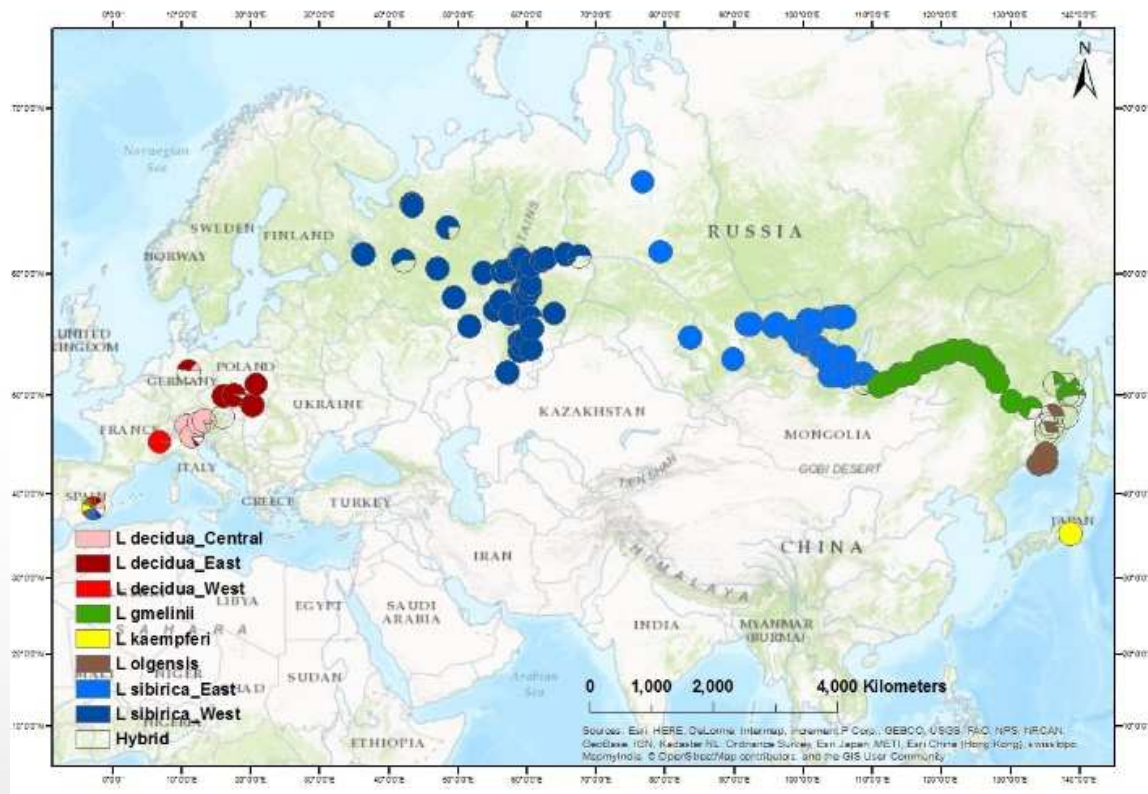


Analysis of SGS disentangle complex situations

- **Tropical species: taxonomy unclear**
- **Species declarations on timber mostly concerns the main well-known species, not all**
 - Cumaru: *Dipteryx odorata*
 - Ipe: *Handroanthus serratifolius/impetiginosus*
 - Jatoba: *Hymenaea courbaril*
- **Set of molecular markers can in some genera both identify species and origin**

Testing species and origin with the same method

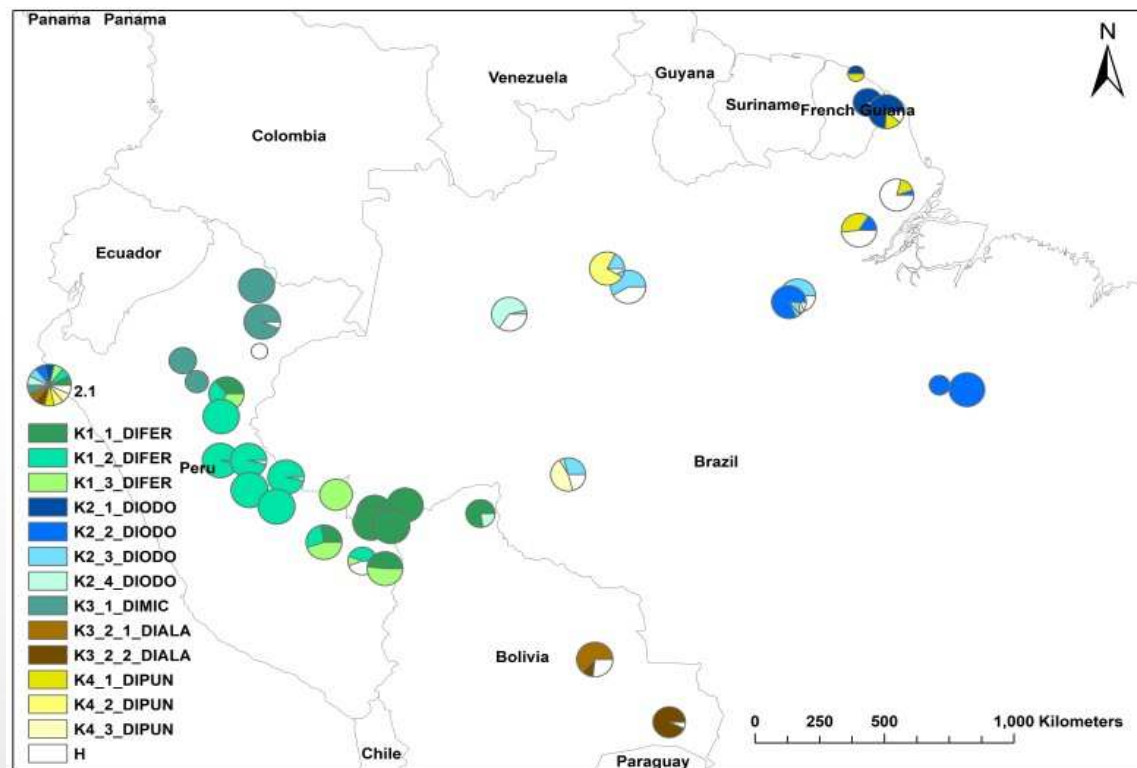
Easy case: species have different distribution ranges



Larix sp.

Testing species and origin with the same method

Easy case: species have different distribution range, **BUT species could be only identified after analysis**



Original Article

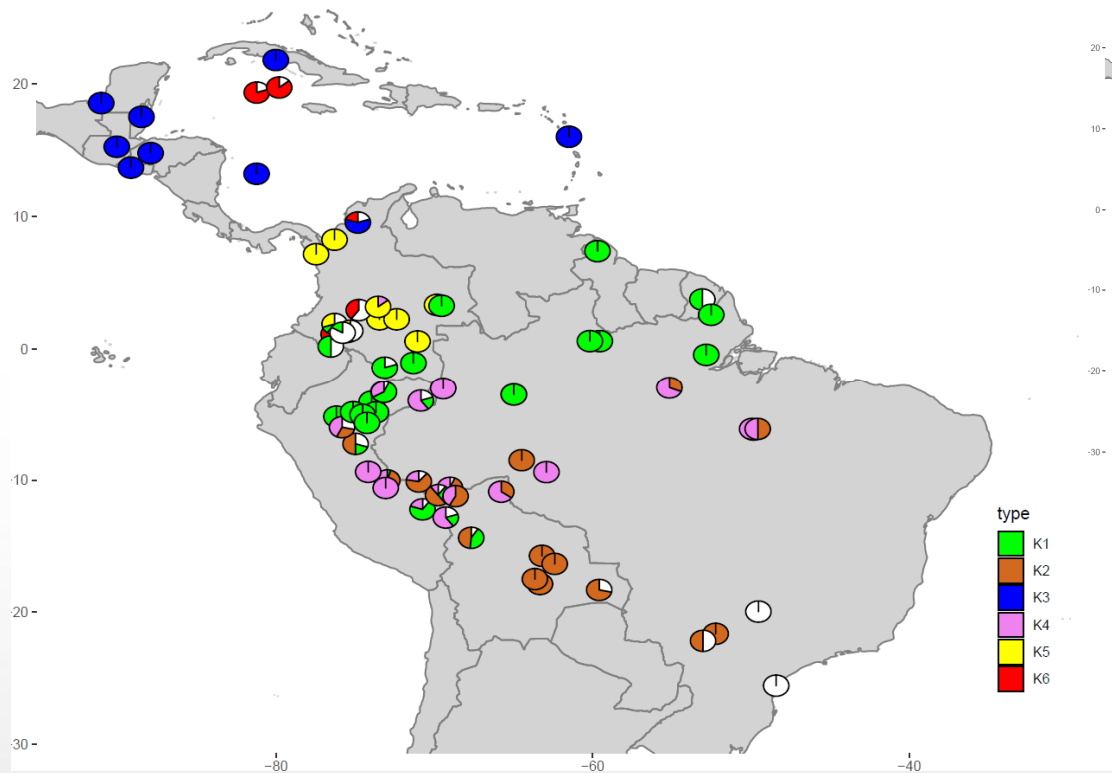
SNP Markers as a Successful Molecular Tool for Assessing Species Identity and Geographic Origin of Trees in the Economically Important South American Legume Genus *Dipterix*

Euridice N. Honorio Coronado, Céline Blanc-Jolivet, Malte Mader, Carmen R. García-Dávila, David Aldana Gomero, Dennis del Castillo Torres, Gerardo Flores Llampazo, Gabriel Hidalgo Pizango, Alexandre M. Sebbenn, Barbara R. V. Meyer-Sand, Kethelyn Paredes-Villanueva, Niklas Trskand, Valerie Troispoux, Marie Massot, Catarina Carvalho, Haroldo C. de Lima, Domingos Cardoso, and Bernd Degen

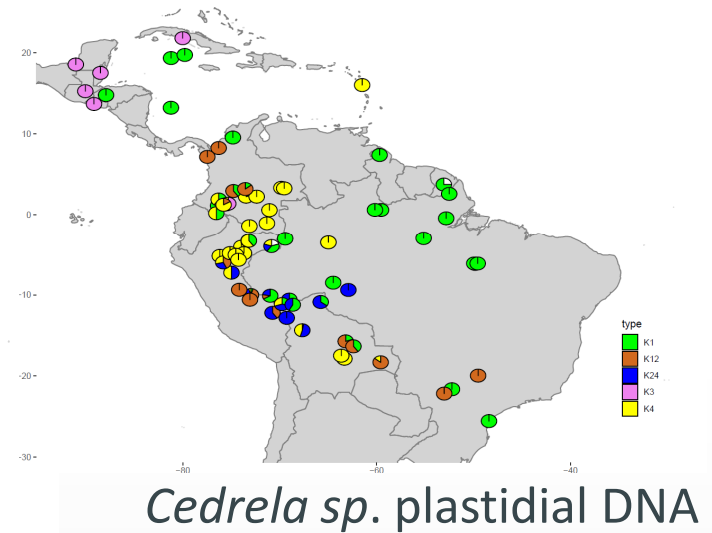
Dipterix sp.

Testing species and origin with the same method

More difficult: some species are **sympatric** in some regions and plastidial DNA gives no species info



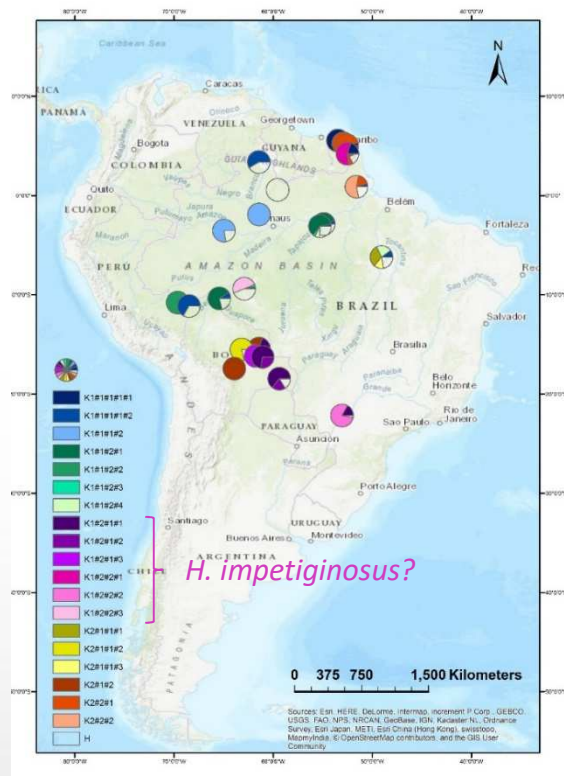
Cedrela odorata, *C. fissilis* (K2), nuclear DNA only



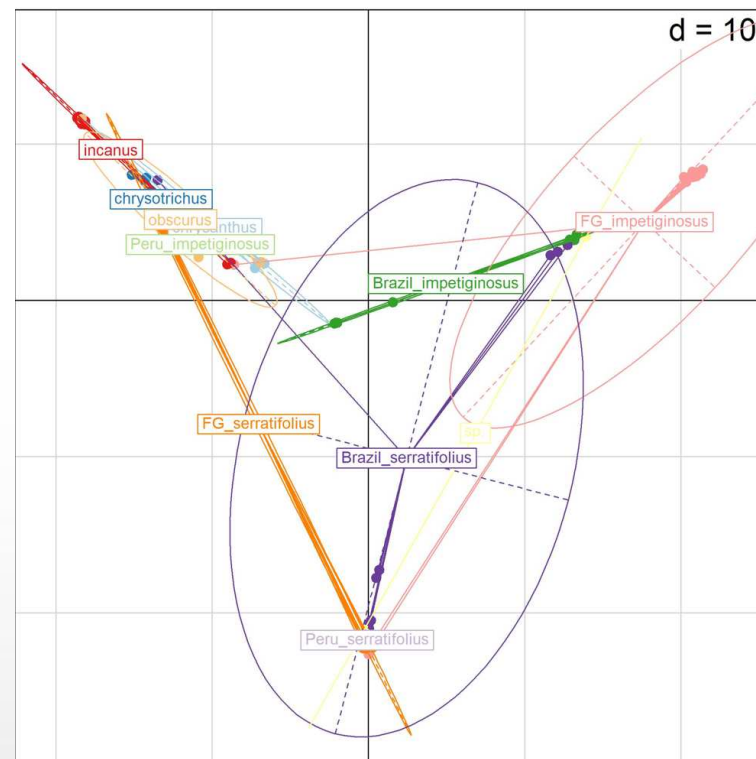
Cedrela sp. plastidial DNA

Testing species and origin with the same method

Challenging case: species are **sympatric AND unclear taxonomy**



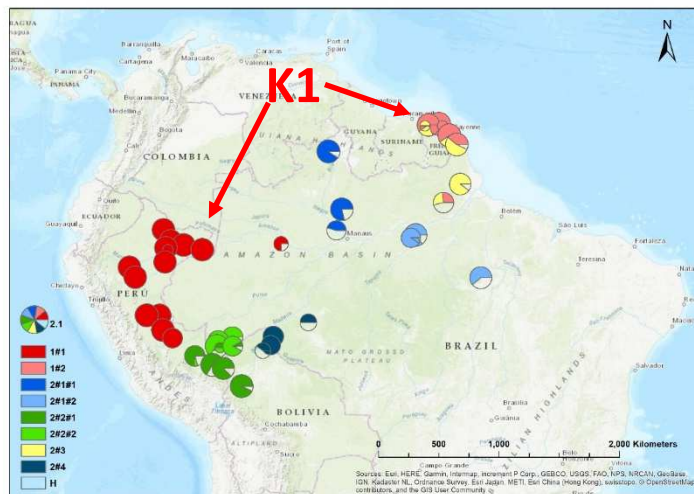
Handroanthus sp. (Nuclear + Plastid)



Handroanthus sp.: species ID loci?

Testing species and origin with the same method

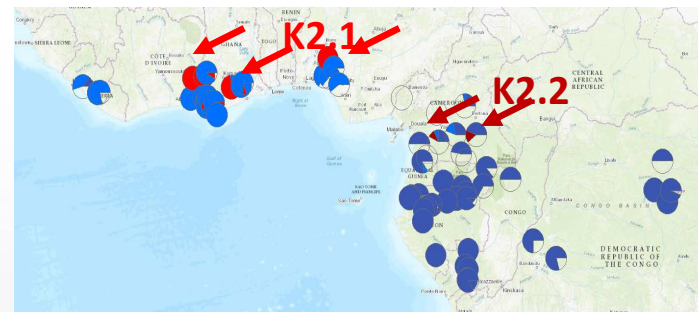
Cryptic species?



Jacaranda copaia (nuclear)



Lophira alata (Nuclear)

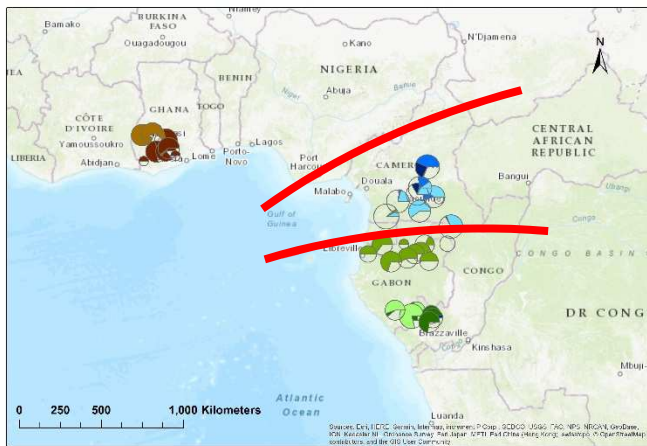


Nauclea diderrichii
(Nuclear)

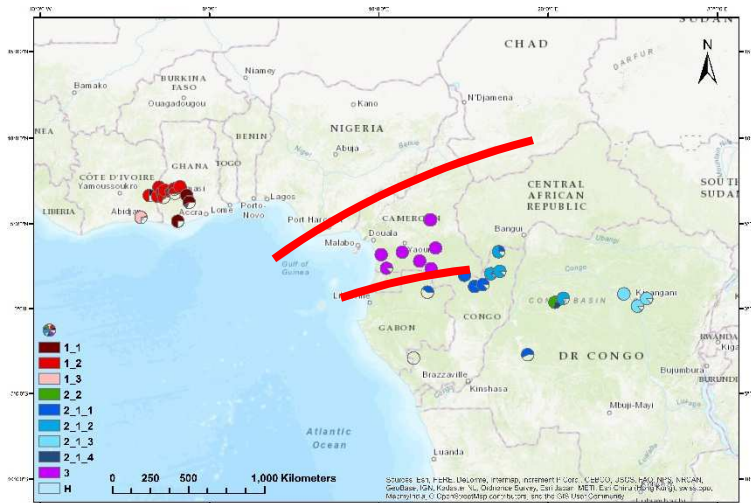
The methods are species-specific... but

- Each species/species complex has a different spatial pattern
 - => separate reference data development
 - => new geographical groups are identified
 - => marker sets are specific
- Some geographical patterns are common among species
 - => evaluate chances of success!!

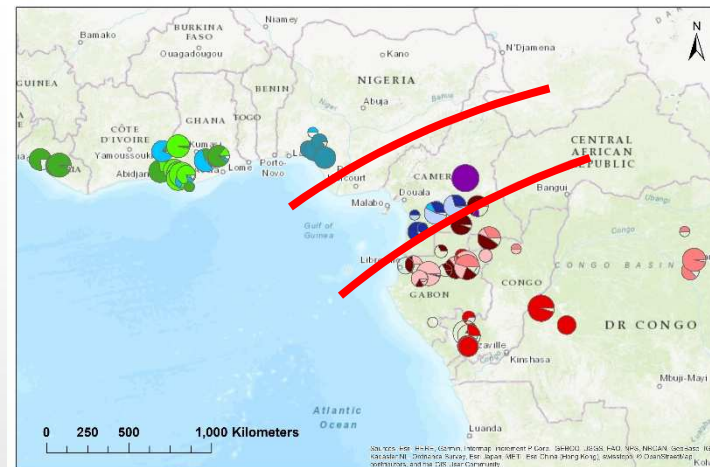
Common patterns among species



Cylicodiscus gabunensis (Nuclear)



Entandrophragma utile (Nuclear+ Plastid)

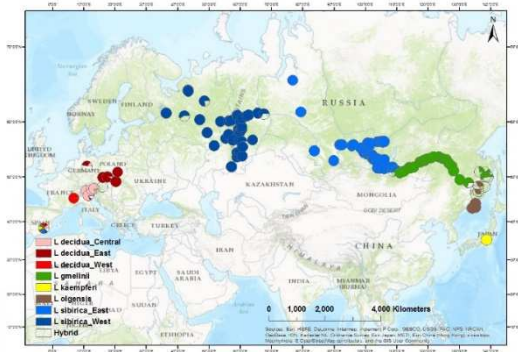


Nauclea diderrichii (Nuclear)

Which steps are necessary to apply the methods?

Testing the reliability of the method (for origin ID)

- Statistical power

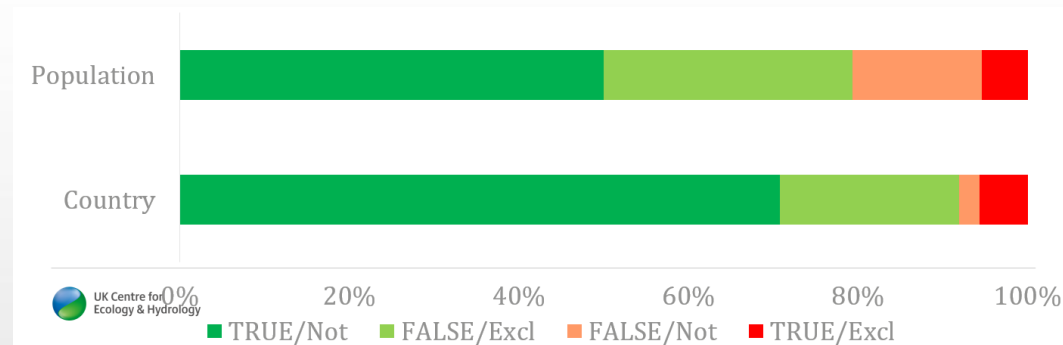


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#####
### Results of Self-Assignment ###
#####
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Group Size = 1
Number of Tests = 1687
Method of assignment = Bayesian Approach
Minimum number of bi-parental loci requested = 1
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Population	Sample Size	Tested ind/groups	% correct assigned
L decidua_West	59	59	46
L decidua_Central	85	85	73
L decidua_East	61	61	49
L sibirica_West	626	626	98
L sibirica_East	503	503	97
L gmelinii	250	250	83
L olgensis	68	68	93
L kaempferi	35	35	100
Total			90
Mean			80

- Blind testing



Thanks !

Hilke Schroeder

Björn Klinkhammer

Stefanie Palczewski

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